87-024752/04 B04 D16 WAKUNAGA SEIYAKU KK

WAKU- 04.06.85 •J6 1280-292-A

B0114

04.06.85-JP-121249 (10.12.86) C12n-15 C12p-21

Protein prepn. by exo-bacterium secretion involves host transformation by introducing recombined DNA into host bacterium cell

C87-010334

USE/ADVANTAGE

B(4-B4A5) D(5-C12)

The protein is obtd. by a simple genetic engineering

EXAMPLE

The vector used is pTA 1529 (1) which is prepd. from pTA 529 and pHS 1.

A gene coding humun-epithelial cell growth factor (II) is combined with (I) to give recombinant DNA (III). E. celi K 12 YK 537 is transformed by (III) to give transformed cells

(IV). (IV) is cultured in LB medium and then in M-9 medium to give a liquid which is then passed through a Prep PAK column and then a DEAE-TOYOPEARL column to collect the desired fraction (II).(18ppW97LDDwgNo0/1).

J61280292-A

A new method for the prepn. of a protein by an extracellular bacterial secretion comprises:

(A) constructing a vector contg. the promoter originated from an alkaline phosphatase gene and a gene coding the signal sequence under the control of this gene, and which can replicate in bacterium host cell;

(B) a gene coding the foreign protein is integrated in to this vector and the recombinant DNA is used to transform the bacterium host cell;

(C) transformed cells are cultured in a medium contg. inorg, phosphorus in amt, insufficient for the induction of protein synthesis and sufficient for the growth of bacteria, and then transferred to a medium to which inorg, phosphorus or a medium contg. it is added at a constant rate; and

(D) the foreign protein is recovered from the cultured liq.

B01 D16

87-024753/04 YAKULT HONSHA KK HONS 06.06.85

•J6 1280-293-A

06.06.85-JP-121488 (10.12.86) C12p-33 C12r-01/64 Steroid phosphoric acid ester prepn. - by microbial conversion using Mortierella fungus

C87-010335

B(1-D1) D(5-C4)

B0115

The process includes a step in which a filamentous fungus belonging to Mortierella species, and able to phosphatise a steroid cpd., is contacted with a steroid cpd. or its alkaline metal salt. Subsequently, the phosphate of the steroid cpd. is recovered.

USE - Prepn. of highly water-sol, steroid cpd. In an example, 61 liq. medium contg. 50 g glucose, 5 g peptone, 2 g yeast extract, 1 g KH2PO4, 2 g K2HPO4, 0.5 g MgSO4.7H2O, 10 mg CaCl2, 10 mg FeSO4.7H2O, 10 mg thiamine-HCl, 1 g taurolithocholic acid and 1 l water is fed into a 10 I fermentor and Y 2-1 species previously cultured in the same medium snnas above at 27 deg.C for 48 hrs. is inoculated into the medium and cultured at 27 deg.C for 5 days with

irring at 300 rpm and aeration of 0.5 vvm (pH: 7-7.5). Then, the cultured liq. is cooled at 50 deg.C and centrifuged to give a clear supernatant liquor. It is passed through an Amberlite XAD-2 column and the absorbed bed is cluated by methanol. The cluate is mixed with an extract of the centrifuged solid and concentrated in vacuo and absorbed on a Sephadex LH20 column and it is eluted by chloroform/methanol and then eluated by methanol, and the latter

water-sol, fraction is conc. in vacuo to give 3.5 g solid. It is purified by a DEAE-Sephadex A-25 column and a XAD-2 column to give 2.1 g of Na taurolith ocholic acid 3 phosphate. (7pp Dwg.No.0/0)

87-024754/04 SANKYO KK

B03 D16

SANY 06.06.85 J6 1280-295-A

06.06.85-JP-121479 (10.12.86) C12p-41 C12r-01/01 Optically active hydroxyethyl azetidinone derivs, prepn. - from inactive acyloxyethyl azetidinone derivs. optically using microorganisms or enzymes

C87-010336

Optically active 8-lactam cpds. (1; Rizil) are produced by selective hydrolysis of racemic cpds. of formula (1) using a microorganism or an enzyme.

$$CH_1 \longrightarrow R_2$$

$$CH_1 \longrightarrow R_3$$

$$R_3 \longrightarrow R_4$$

$$(1)$$

R₁ = opt. substd. ncyl;

R, = opt. substd. alkyl. alkenyl, alkynyl, aryl, alkylthio, alkylsulphonyl, arylthio or arylsulphonyl or acyloxy; and

R, = It or protective gp. for N atom.

USE/ADVANTAGE

Optically active 3-(1-hydroxyethyl)-2-azetidinone deriv.

B(7-D1) D(5-C)

B0116

is obtd. from optically inactive 3-(1-acyloxycthyl)-2-azetidinone deriv.

These optically active azetidinone derivs, are important intermediates for carbapenem and penem derivs, which have antibacterial activity.

MICROORGANISM

This may be chosen from bacteria, yeast and fungi: Dacteria:

Arthrobacter simplex SANK 73560 (IAM 1660);

Chromobacterium violaceum SANK 72783 (ATCC 31532): Flavobacterium capsulatum SANK 70979 (IFO 12533);

Flavobacterium meningosepticum SANK 70779 (1FO 12535); or Bacillus subtilis SANK 76759 (IAM 1069);

Yeast:

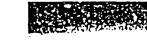
Aureobacidium pullulans SANK 10877 (ATCC 15232): Candida albienus SANK 50169 (ITO 0683):

Pichia farinosa SANK 58062 (LAM 4303): Pichia terricola SANK 51684 (FERM 8001):

Rhodotorula minuta SANK 50871 (HO 0932); or Saccharomyces cerevisine SANK 50161(IAM 4512);

Fungi:

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Aspergillus niger SANK 13658 (ATCC 9142); Gliocladium roseum SANK 10560 (FERM 8259); or Humicola asteroidea SANK 14981 (FERM 8260).

This may be of microorganism or animal or plant cell origin, examples of which are: esterase (carboxylic-ester hydrolase, EC 3.1.1.1, e.g. pig

liver originated commercial prod. PLE);

lipase (trincylglycerol ncylhydrolase, EC 3.1.1.3, e.g. Aspergillus oryzne or Aspergillus niger-originated commercial prod.):

aminoacylase (N-Amino acid aminohydrolase, EC 3.5.1.14 e.g. commercial prod. prepd. from Aspergillus genus of fungi).

Commercially available low-cost crude prod, such as Takadiastase (originated from Aspergillus oryzae) contains lipase and may be used in place of purified standard lipase.

dl-3,4-Trans-1-(4-methoxyphenyl)-3-[(IR*)-l-neet xyechyl]--4-ethynyl-2-azetidinone (60 mg) was subjected to skaken culture with Pichia farinosa SANK 58062 (IAM 4303) at 30°C for 24 hrs.

Culture liquor was extracted with ethyl acetate, and obtd. crude prod. (76 mg) was purified by silica gel TLC (cyclo-hexane/ethyl acetate = 1/1, U.V. lamp detection, Rf = 0.32) to give (21 mg) of (38,48)-1-)4-Methoxyphenyl)-3-((1R)-1-hydroxyphenyl)-3-((1R -hydroxyethyl)-4-ethynyl-2-azetidinone. $(a)_{11}^{24} = -135^{\circ}$ (C=1, CI:Cl;).(22ppW-69LDDwgNo.0/0).

JG1280295-A

87-024755/04 B05 D16 SUMITOMO CHEM IND KK

SUMO 05.06.85 J6 1280-296-A

B(10-E4B) D(5-A2C)

B0117

05.06.85-JP-121944 (10.12.86) C12p-41 Biochemical prepn. of optically active phenoxy phenoxy proponol involves reacting bacterial esterase with opt, said, organic carboxylic ocid ester C87-010337

Optical biochemical resolution of (++) -2-(4-phenoxyphenoxy) proponed of (1) comprises interacting esterase produced by microorganism selected from the gp. consisting of Pseudomonas, Chromobacterium, Arthrobacter, Alcaligenes, Candida, Arthrobacter, Tolulopsis, Achromobacter, Nocardia, Flavobacterium, Achromobacter, Nocardia, Flavobacterium, Tolulopsis, Brevibacterium, Bacilius, Escherichia, Micrococcus, Hansenula, Mucor, Corynebacterium, Mycobacterium, Saccharomyces, Thermomyces, Humicola, Thizopus, Aspergillus, Streptomyces, Geotricum, Treoderma, Acinetobacter, Aeroinonas, Beauveria, Rhodotorula, Enterobacter, Penicillium, Serratia, Erwinia, Staphylococcus, Phycomyces, Propionibacterium, Metarrhizium, Necardia, Saccharomyces, Verticillium, and Anniones. Pacecilomyces, Saccharomycopsis, Verticillium and X anthomonas, with organic 1-18C opt. said. carboxylic acid ester of (+-)-(1) to resolve to optically active (1) and its antipode ester.

Cultivation is conducted at 20-40 deg.C for 1-3 days in liq. medium. As esterase there are used culture liquid, cells septi. from the culture liq., crude esterase sepd. from the cells or culture filtrate, culture filtrate contg. esterase, purified esterase and esterase-contg. extract

of concentrate. Reaction is conducted under sliaking or stirring. The reaction temp. is 10-70 deg.C. To keep the pli constant during the reaction, buffer such as sodium phosphate and sodium acetate can be used. Use concn. of the substrate is 0.5-80 wt.%, pref. 10-50 wt.%. Pref. 2-12C organic carboxylic acid is used.

ADVANTAGE - Process gives optically active (1) with very high optical purity. (11pp Dwg.No.0/0)

87-024756/04 BO4 D16 S03 (D13) NODA INST SCI RES

NODA 04.06.85 *J6 1280-297-A

04.06.85-JP-119782 (10.12.86) C12q-01/26 G01n-33/50 Determin, of amadori cpd. in e.g. soy sauce - by treatment with fructosyl:amino acid oxidase and e.g. determn, of hydrogen peroxide

C87-010338

Determination of Amadori end, comprises treating a 11q. contg. Amadori cpd, with fructosylamino acid oxidase in the presence of oxygen, and determg, the amt, of oxygen consumed in the oxidn. reaction or determs, hydrogen peroxide formed by the reaction.

Reagent for the determn, of Amadori cpd, contains fructosylamino acid oxidase.

Amadori cpd. is that formed from aldose and alpha-amino acid, from glucose and alanine fructosylalanine hydroxyacetonyiglycine from glyceraldehyde and glycine. Sample liq. contg. Amadori cpd. is e.g. soy sauce, honey, etc. Fructosylainino acid oxidase used is pref. that obtd. by cultivating microorganism, esp. bacteria belonging to Corynebacterium genus (e.g. Corynebacterium sp. No.2-3-1). The determin of oxygen is carried out by oxygen electrode, and that of hydrogen peroxide by

colorimetry.

ADVANTAGE - The determin, of Amadori cpd. can be easily carried out. Amadori cpd. reflects the state of food (e.g. soy sauce) or infusion liq. during mfr. or storage. Amadori cpd. bound by B(4-B2C2, 4-B4B, 4-B4D5, 4-D1, 7-A2, 10-B2J, 11-C7B1, 12-K4A) D(5-A2A) 80118

protein can be determed. after conversion into its free state by the reaction with a suitable peptidase. This is useful for the examination of diabetes mellitus, (8pp Dwg.No.0/3)

